

## Application note No. 214

# NucleoCounter® NC-200™

## Viability and Cell Count using the Via1-Cassette™ with Reagent A100 and B

### Product description

The NucleoCounter® NC-200™ system enables the user to perform automated cell counting of a broad range of mammalian.

### Application

The Via1-Cassette™, **Reagent A100** and **B** used together with the NucleoCounter® NC-200™ can be used for determining of the viability and cell concentration of aggregating cells. Treatment of cell samples with **Reagent A100** facilitates lysis and thereby disaggregation of cell aggregates resulting in single cell nuclei suspensions. **Reagent A100** also enables staining of all cells with DAPI. **Reagent B** stabilizes the nuclei for the analysis.

It is recommended that the user do a complete validation of this assay before implementing, since the validation done by ChemoMetec A/S has been limited due to the

amount of variation in cell types, aggregation level and surfaces the cells adhere to, e.g. micro carrier.

### Introduction

In order to determine the cell concentration, a sample containing cells in suspension is diluted with **Reagent A100** (lysis buffer) followed by stabilization with **Reagent B** and drawn into the Via1-Cassette™. The inside of the Via1-Cassette™ is coated with DAPI, which after lysis with **Reagent A100** stains all cell nuclei in the sample. When the cell sample is not treated with **Reagent A100**, the dye only stains the non-viable cells. The volume of each Via1-Cassette™ has been calibrated to give high precision in the resulting count.

The Via1-Cassette™ is placed in the NucleoCounter® NC-200™ where cell concentration is determined.

### Procedures

If the cell line to be investigated is adherent or semi-adherent, then start by getting all cells into suspension using the preferred method of your laboratory (e.g. trypsin/EDTA treatment).

### Materials needed

- Cells to be counted
- **Reagent A100**
- **Reagent B**
- Two Via1-Cassettes™

1. The first step is to determine the total cell concentration of the cell sample.
  - a. The cell suspension is mixed to obtain a homogenous suspension and a sample is mixed 1:1 with **Reagent A100**. For example, to 100 µl of cell suspension add 100 µl of **Reagent A100** and mix by pipetting.
  - b. Add one volume of **Reagent B** to the mixture of cell suspension and **Reagent A100**. Using the example above, to 200 µl of the mixture of cell suspension and **Reagent A100** add 100 µl of **Reagent B** and mix by pipetting.
  - c. Draw a cell sample into the first Via1-Cassette™ by inserting the tip into the diluted cell suspension and pressing the piston.
  - d. Select the “**Viability and Cell Count – A100 and B Assay**” and press RUN. Click “OK” when the loaded Via1-Cassette™ containing the sample diluted 1:1:1 with **Reagent A100** and **B** is in place in the NucleoCounter® NC-200™.

2. The second step is to determine the concentration of non-viable cells of an undiluted cell sample.
  - a. The undiluted cell suspension (without **Reagent A100** and **B** treatment) is mixed again to obtain a homogenous suspension. Draw a cell sample into the second Via1-Cassette™ by inserting the tip into the cell suspension and pressing the piston.
  - b. Follow the on-screen prompts instructing to replace the first Via1-Cassette™ with the second Via1-Cassette™ loaded with the undiluted cell suspension and click “OK”.

After approximately 2 minutes the cell concentration (cells/ml) of the total cell count and the viability are displayed together with the cell diameter. The displayed cell concentration of the total cell count has been compensated for the dilution caused by the addition of **Reagent A100** and **B**. If the cell sample has been further diluted or concentrated and the user has entered the volumes into the user interface the dilution factor has also been taken into account and the returned cell concentration is for the original cell concentration.

### Note

To assure reliable results, it is recommended that the cell concentration of the counted cell suspension should be in the range of  $5 \cdot 10^4$  cells/ml to  $5 \cdot 10^6$  cells/ml. If the cell concentration is above  $5 \cdot 10^6$  cells/ml, the cell suspension can be diluted with growth media or PBS to achieve the desired concentration. The diluted cell sample is then treated as described above. By inserting the value for the dilution volume in the dilution field on the user interface the returned cell concentration is for the original cell sample.

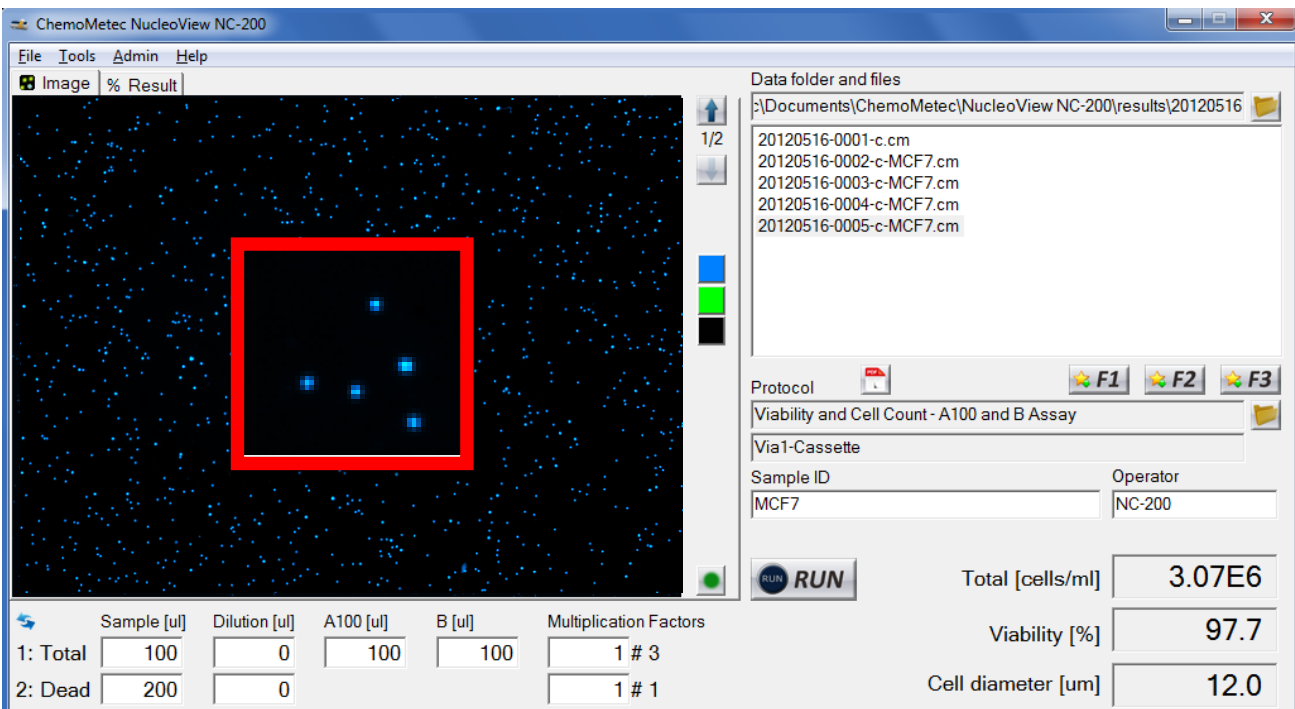
If the concentration of cells is below  $5 \cdot 10^4$  cells/ml then the cell concentration may be increased by centrifugation followed by resuspension of the pellet using growth media or PBS. The resuspended cell sample is then treated as described in the procedure. By inserting a negative value representing the volume removed from the sample in the dilution field on the user interface the returned cell concentration is for the original cell sample.

### Viability

The viability is calculated as follows:

$$\% \text{ viability} = \frac{C_t - C_{nv}}{C_t} \cdot 100\%$$

<b>% viability</b>	The percentage of viable cells in the cell suspension
<b>C<sub>t</sub></b>	The total concentration of cells
<b>C<sub>nv</sub></b>	The concentration of non-viable cells



The screenshot shows the ChemoMetec NucleoView NC-200 software interface. The main window displays a field of blue fluorescent cells. A red square highlights a specific region. On the right, a file list shows several .cm files. Below the image, a table provides assay parameters and results.

	Sample [ul]	Dilution [ul]	A100 [ul]	B [ul]	Multiplication Factors
1: Total	100	0	100	100	1 # 3
2: Dead	200	0			1 # 1

Total [cells/ml]	3.07E6
Viability [%]	97.7
Cell diameter [um]	12.0

Determination of the cell concentration of aggregated MCF7 cells. The cells for the first Via1-Cassette™ were disaggregated by adding **Reagent A100** and **B** and the cells for the second Via1-Cassette™ were untreated. The two samples were analyzed using the Viability and Cell Count – A100 and B Assay. The total cell population is stained with DAPI and appears blue. An insert shows a close up of parts of the images. The results are presented at the bottom right and extended results are presented in the result tab page.

## Troubleshooting

### Inaccurate and imprecise counting:

When setting up a new cell line it is important to inspect that the cell line is counted correctly. The cells included in the total count can be marked by clicking on the overlay button in the bottom right corner of the image. Visual inspect the image to evaluate in the vast majority of the cells has been counted correctly. If this is not the case right click on the image file in question and choose “Show Counting Gates in Plot Manager”. Inspect the gates displayed in the Plot Manager. If the gating is inappropriate adapt the gate(s) to cover the cell population (do not include debris and very large objects) using the Protocol Adaptation Wizard and save the changes to a new protocol. Note that the user is responsible for defining appropriate gating of the particular cell line.

### Warning that the cell concentration of non-viable cells is higher than the total cell concentration:

Make sure the problem is not due to interchanged samples of the total count sample treated with **Reagent A100** and **B** and the non-viable count sample without treatment. If the samples have not been interchanged, the continued warning can be due to a very high frequency of non-viable cells in the sample.

#### Handling and storage

For handling and storage of ChemoMetec instruments and reagents, cassettes and NC-slides refer to the corresponding product documentation. For other reagents refer to the material data sheet from the manufacturer of the reagents and chemicals.

#### Warnings and precautions

For safe handling and disposal of the ChemoMetec reagents, cassettes and NC-slides refer to the corresponding product documentation and the NucleoCounter® NC-200™ user's guide. For other reagents refer to the safety data sheet from the manufacturer of the reagents and chemicals required for this protocol. Wear suitable eye protection and protective clothes and gloves when handling biologically active materials.

#### Limitations

The NucleoCounter® NC-200™ system is FOR RESEARCH USE ONLY. NOT FOR DIAGNOSTIC OR THERAPEUTIC USE. The results presented by the NucleoCounter® NC-200™ system depend on correct use of the reagents, Cassettes and the NucleoCounter® NC-200™ instrument and might depend on the type of cells being analyzed. Refer to the NucleoCounter® NC-200™ user's guide for instructions and limitations.

#### Liability disclaimer

This application note is for RESEARCH PURPOSES ONLY. It is not intended for food, drug, household, or cosmetic use. Its use must be supervised by a technically qualified individual experienced in handling potentially hazardous chemicals. The above information is correct to the best of our knowledge. Users should make independent decisions regarding completeness of the information based on all sources available. ChemoMetec A/S shall not be held liable for any damage resulting from handling or contact with the above product.

#### Product disclaimer

ChemoMetec A/S reserves the right to introduce changes in the product to incorporate new technology. This application note is subject to change without notice.

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